



Phytochemistry analysis and modulatory activity of *Portulaca oleracea* and *Aquilaria malaccensis* extracts against High-fructose and high-fat diet induced immune cells alteration and heart lipid peroxidation in Rats

Samir Derouiche*, Ouidad Degachi and Khaoula Gharbi

Department of Cellular and Molecular Biology, Faculty of natural sciences and life, University of ElOued, El-Oued 39000, Algeria
dersamebio@gmail.com

Available online at: www.isca.in, www.isca.me

Received 9th September 2018, revised 25th February 2019, accepted 30th March 2019

Abstract

Our objective of this investigation is to estimate the influence of *Portulaca oleracea* (*P. oleracea*) and *Aquilaria malaccensis* (*A. malaccensis*) methanol extracts on High-fructose-fat diet (HFFD) induced immune cells alteration and heart lipid peroxidation in Rats. Twenty five Females rats were equally divided into five groups (n=5) as control, HFFD, HFFD+Po, HFFD+Am and HFFD+Po+Am groups. High fructose-fat diet was added in diet of rats with (60% fructose and 60% kcal fat) for 70 days. Methanol extracts of *P. oleracea* (Po) (400mg/kg bw) and *A. malaccensis* (Am) (200mg/kg bw) were supplemented orally for four weeks. Methanol extracts of plants were prepared and phytochemicals were analyzed by using HPLC methods. Hematological markers and lipid peroxidation in heart were assessed. Results obtained shown that HFFD induction caused a significant increase in White blood cell ($P<0.01$), Granulocyte ($P<0.05$), Lymphocyte ($P<0.01$) Monocyte ($P<0.001$) count and heart MDA level and no significant effect in Red blood cell and Hemoglobin level compared to the rats given normal diet. Methanol extracts of *P. oleracea* and *A. malaccensis* treatment partially correct the parameters studied. Our study indicate that *A. malaccensis* possesses the ability to control the heart lipid peroxidation and immune cells alteration associated with High fructose-fat diet.

Keywords: *P. oleracea*, *A. malaccensis*, lipid peroxidation, immune cells, fructose, high fat diet.

Introduction

Nutrition is an essential element of life which is directly related to the health of the population; Therefore, healthy nutrition is a concern of consumers and nutritionists¹. A food imbalance is a major cause for various pathologies such as diabetes and obesity². Higher levels of fat in foods increase the levels of lipids, such as cholesterol in the blood, increasing the risk of atherosclerosis, obesity, hypertension, diabetes and breast cancer³. On the other hand, in an experimental study, food rich in fructose (more than 60% of diet) for 6 months causes the initiation or progression of many diseases such as diabetes and dyslipidemia in rats⁴. In addition, a high calorie diet may cause the risk of liver insulin resistance, a build-up of fat in the liver and skeletal muscle⁵. Oxidative stress is an important factor causing metabolic and physiological alterations and various diseases in the body⁶. Since long time, plants have been used as medicine against several diseases for many diseases; they are still the basis of a system of traditional medicine in different cultures⁷. Among these medicinal plants is *Portulaca oleracea* which is a traditional vegetable very used by indigenous and tribal peoples in many countries. It is known to contain many active substances are also considered as sources of many dietary supplements⁸. On the other hand *Aquilaria malaccensis* is a plant of the family Thymelaeaceae belongs to the tropical species⁹, is one of the main sources of agarwood, which

provides clues about their pharmacological properties. Indeed, agar wood is very rich in bioactive substances which favor their use in traditional medicine¹⁰. In light of these information, our objective for this work is to estimate the benefic role of methanol extracts of *Portulaca oleracea* leaves and the trunk bark of *Aquilaria malaccensis* on high fructose-fat diet induced immune cells disturbance and heart lipid peroxidation in rats.

Materials and methods

Chemicals: Fructose was used as powder provide from (Biomax), (Specialized Food Industry, Algiers, Algeria). All other chemicals are of fine analytical grade.

Plant material: The plants used in this study are the bark trunk *Aquilaria malaccensis* (*A. malaccensis*) were purchased from the local market and the leaves of *Portulaca oleracea* (*P. oleracea*) was harvested in the region of El-Oued "Guemar" in September 2016. The bark of *A. malaccensis* and leave of *P. oleracea* were washed and dried in an airy place and then crushed and stored in powder form until the beginning of the experiment.

Preparation of Methanol extracts: The crude samples (each 3.75g) of each powder mixed with 25ml of methanol-water (70:30) at 25°C for 48h and then filtering with filter paper. After

extraction, the solvents were removed under low temperature at 40°C using a rotary evaporator.

Method of Phytochemistry HPLC analysis: The methanol extracts of *Portulaca oleracea* and *Aquilaria malaccensis* were filtered before injection. An HPLC system was used with a detector at $\lambda=280\text{nm}$ for polyphenols and 360nm for flavonoids, the experimental conditions are as follows:

The column used is 150mmx4.6mm with C18 as stationary phase, Mobile phase: acetonitrile and glacial acetic acid (2%, pH=2.6 (30°C)). The identification of the peaks of polyphenols and flavonoids was made thanks to the standard achieved by pure components by comparing the retention times.

Animal study: Animals and treatment: Adult female albino rats (aged between 8 to 10 weeks), weighing 200–270g, rats were grouped into five lots of 5 rats in each and reserved in the pet store of our institute. After the adaptation period for two weeks under laboratory conditions (humidity 64.5%, $T^{\circ}=25\pm 2^{\circ}\text{C}$ and photoperiod (light/dark) 12h/12 h, Put water and food at the disposal of rats freely. Food is rich on fructose (60% of fructose of diet)¹¹ and fat diet (60% fat)¹² (HFFD) ad libitum for 70 days. The five groups of rats are distributed as follows:

Group 1 (control group): animals were given normal diet served as control.

Group 2 (HFFD): Rats were given HFFD diet.

Group 3 (HFFD+Po): Rats were given HFFD diet plus methanol extracts of *P.oleracea* (400 mg kg⁻¹ d⁻¹) administered orally.

Group 4 (HFFD+Am): Rats were given HFFD diet plus methanol extracts of *A.malaccensis* (200mg.kg⁻¹.d⁻¹) administered orally.

Group 5 (HFFD+Po+Am): Rats were given HFFD diet plus both methanol extracts of *P.oleracea* and *A.malaccensis* administered orally.

The duration of different treatment is 30 days. Body weight was recorded regularly.

Collection of biological materials: After 30 days of treatment, the rats are sacrificed after at least 16 hours of fasting by decapitation under inhalation chloroform anesthetized. The sample of blood is carried in tubes of EDTA for hematological studies (hematological analysis (FNS) is performed by the hematology auto analyzer (Sysmex)). Heart was quickly removed, absolute heart weight was determined than stored at -20°C for lipid peroxidation analysis.

Lipid peroxidation measurement: Preparation of homogenates: Mixt 1g of heart tissue with 9ml of buffer solution (KH₂PO₄, pH=7.4). After grinding of the mixture, the homogenate is obtained by used centrifugation at 9000xg, 15min at 4°C than the homogenate samples are stored at -20°C for MDA assay.

Estimation of Malondialdehyde (MDA) levels: Malondialdehyde analysis is based on the thiobarbituric acid (TBA) technique following the method of Sastre et al.¹³ by measuring the absorbance of TBA-MDA complex at 530nm. The results presented by $\mu\text{Mol/mg}$ protein.

Protein determination: Estimation of the tissue protein is based on the Bradford method¹⁴. The standard curve is made by the BSA.

Statistical Analysis: The present results were reported as Mean and standard error of mean. The comparison between the means is made by Student t test. Statistically significance threshold of results using $\alpha=p<0.05$. * $p<0.05$, ** $p<0.01$, *** $p<0.001$: comparison with control group, a $p<0.05$, bp <0.01 , cp <0.001 : comparison with HFFD group.

Results and discussion

Polyphenol HPLC analyzes: Chromatogram of HPLC analysis (Figure-1) show that the methanol extract of *P. oleracia* and *A. malaccensis* are rich in polyphenol in different quantities and that Apigenin, Epicatechin and Naringenin has the major substance among these constituents of *P.oleracia* and *A.malaccensis*.

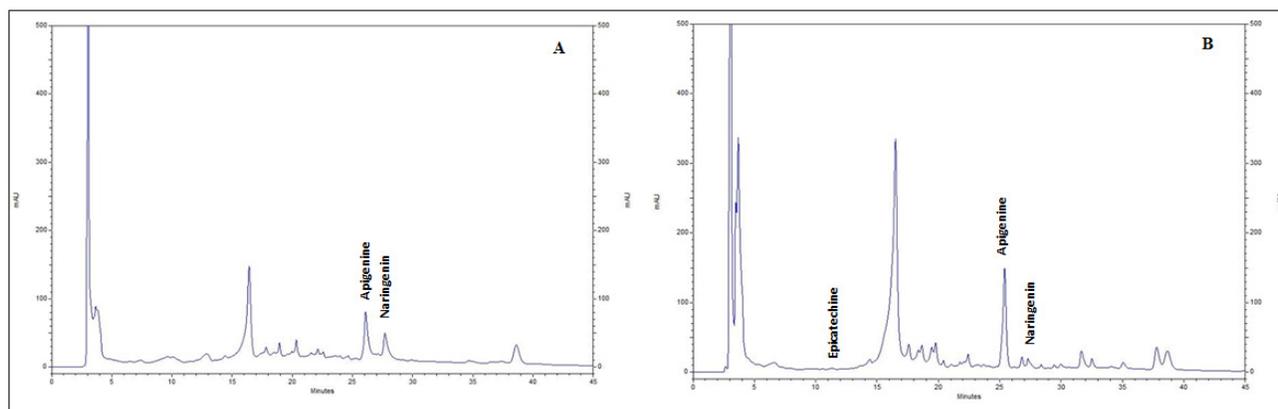


Figure-1: Chromatogram of polyphenol for the methanol extract of *P.oleracia* (A) and *A.malaccensis* (B).

Flavonoid HPLC analyzes: The results of the chromatographic analysis by HPLC show that the methanol extract of *P.oleracia* and *A.malaccensis* are rich in flavonoids in different quantities and that Quercetin, Kampferol and Rutin has the majority substance for *P.oleracia* and *A.malaccensis* (Figure-2).

Body weight and relative heart weight: As seen in Table-1, results show that compared to the control, the weight gain was increased ($p<0.01$) in group of rats under HFFD. However, methanol extract treatment causes a significant decrease ($p<0.05$, $p<0.001$ and $p<0.01$) of weight gain in HFFD+Po, HFFD+Am and HFFD+Am+Po groups respectively compared to the HFFD group. The results obtained show a no significant variation in the relative weight of heart between HFFD and control groups (Table-1).

Hematological markers: As seen from Table-2, results showed a significant increase of White blood cell (WBC) ($P<0.01$),

Lymphocyte (LYM) ($P<0.01$), Granulocyte (GRN) ($P<0.05$) and Monocyte (MON) ($P<0.001$) count and no significant change of RBC and Hemoglobin (HGB) level in HFFD animals as compared to normal animals. Moreover, the results obtained show that there is a significant improvement in the WBC, LYM, GRN and MON count in the rats treated with extract of *A. malaccensis* (Am) and Po+Am compared to HFFD group. While, only *P. oleracea* (Po) treatment significantly decreases the WBC count compared to the HFFD group.

Level of lipid peroxidation marker: The obtained results show that compared to control, the lipid peroxidation in heart was increased ($p<0.05$) in HFFD group. While treatment with *A.malaccensis* (Am) or Po+Am leads to reduction ($p<0.05$) in heart MDA levels when compared to the HFFD group. But no effect of *P. oleracea* (Po) extract treatment on heart lipid peroxidation compared to HFFD group (Figure-3).

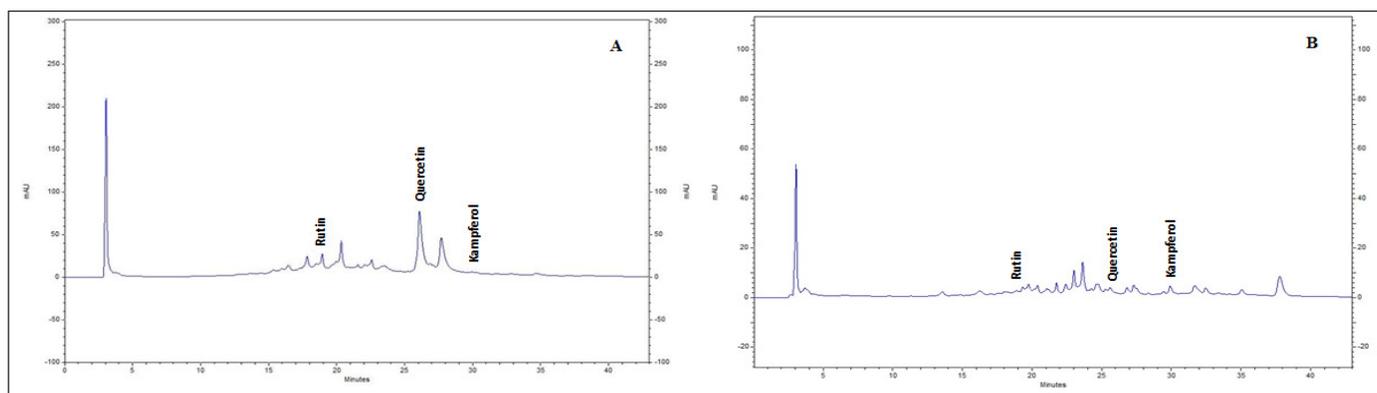


Figure-2: Chromatogram of flavonoids for the methanol extract of *P. oleracia* (A) and *A. malaccensis* (B).

Table-1: Body weight and relative heart weight of experimental groups.

Groups	Control	HFFD	HFFD + Po	HFFD +Am	HFFD +Am+Po
Initial body weight (g)	203.3±2.3	268.5±1.96	239.25±6.87	241±4.04	230.75±5.06
Body weight gain (g/day)	0.53±0.055	1.05±0.036**	0.26±0.25 ^a	0.14±0.06 ^c	0.8±0.06 ^b
Relative heart Weight (g/100g b.w)	2.8±0.05	2.7±0.16	2.9±0.20	2.9±0.09	2.8±0.16

Table-2: Meanhematological markers in experimental groups.

Groups	Control	HFFD	HFFD + Po	HFFD +Am	HFFD +Am+Po
RBC (10 ⁶ /μL)	7.02±0.20	6.86±0.22	6.95±0.15	7.22±0.23	7.13±0.11
HGB (g/dL)	13.42±0.36	13.03±0.37	13.37±0.33	13.85±0.45	13.55±0.42
WBC (10 ³ /uL)	3.62±0.14	4.9±0.12**	4.37±0.18*	4.0±0.11 ^a	3.27±0.27 ^a
LYM (U/l)	3.08±0.12	4.03±0.09**	3.98±0.05* ^a	3.68±0.1 ^a	2.86±0.12 ^b
MON (10 ³ /μl)	0.28±0.01	0.68±0.01***	0.57±0.06*	0.45±0.05* ^a	0.35±0.05 ^b
GRN (10 ³ /μl)	0.34±0.04	0.53±0.02**	0.46±0.01	0.39±0.06 ^a	0.31±0.01 ^a

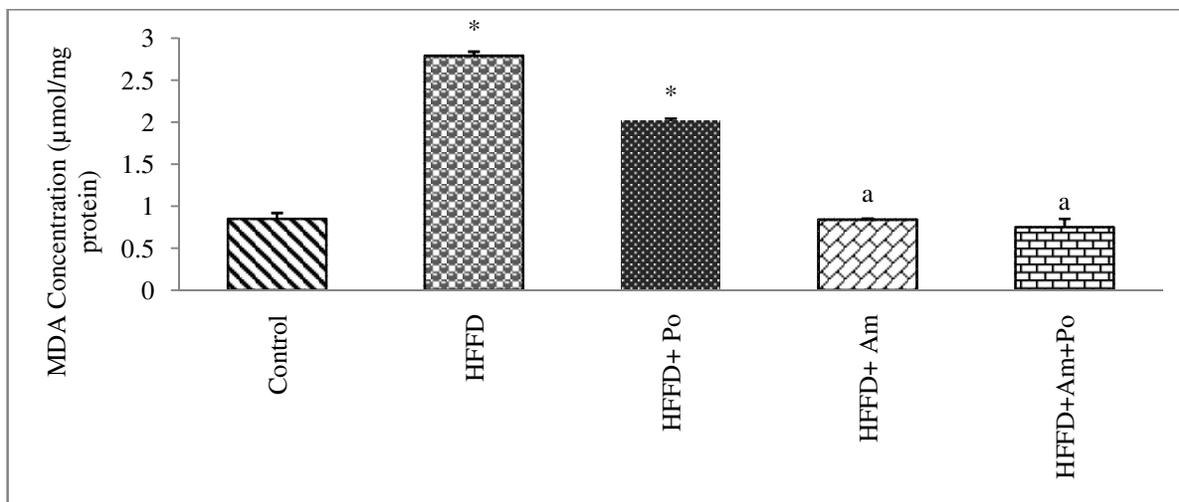


Figure-3: Mean MDA level in heart tissue of control and experimental rats.

Discussion: The results of chromatography HPLC analysis reveal that the methanol extract of dry leaves of *Portulaca oleracea* and the trunk bark of *Aquilaria malaccensis* contain several bioactive compounds, including Apigenin, Epicatechin, Naringenin Quercetin, Kampferol and Rutin. Phytochemicals in the diet can exert on different targets that can relieve multiple pathological processes, including oxidative damage, epigenetic alterations, chronic inflammation, active stimulators, inhibitors and growth terminators and prevention of various diseases associated with oxidative stress^{15,16}. In our study, results show that HFFD food causes a rise in weight gain resulting an obese phenotype of rats. More consumption of high energy content nutrients such as fructose and HFD leads to an increase in the fat mass and fat cell expansion (hypertrophy) without changing food intake, producing the specific pathology of obesity¹⁷. Also high proportion of lipids in food can increase palatability and cause hyperphagia of animals leads to rapid weight gain¹⁸. Treatment of HFFD rats with the methanol extract of *P.oleracea* and / or *A.malaccensis* induces a decrease in body weight gain. The anti-obesity properties of plants can be exerted according to different modes of action: by direct effect on food intake by suppressing appetite and inducing the feeling of satiety, a reduction of lipid absorption, a reduction of energy consumption, increase in energy consumption, slow development of pre-adipocytes, decrease energy intake from the gastrointestinal tract^{19,20}. Our results reveal that the administration of HFFD in rats causes an increase in white blood cells, granulocytes, as well as lymphocytes and monocytes compared to control, these results are of support with study of Gomez-Smith et al.²¹ who used high fat diet model alone and the Pektas et al. study, which showed that levels of pro-inflammatory factors increased by high fructose content²². A high fructose diet may induce inflammation, as fructose in combination with an increase in monocyte-macrophage infiltration²³. In addition, fructose exerts a pro-oxidative and pro-inflammatory effect, since it activates a pro-oxidant enzyme NADPH-oxidase, responsible for the release of superoxide anion²⁴. White blood cell and lymphocyte count analysis is a

very important diagnostic test to detect the presence of inflammation and other diseases²⁵. A complete blood count also is needed to control the meaning of the changes of the immune cells in the body, such as, the elevation of the granulocyte presents a risk of attacking a heart failure in the human, whereas a decreased risk has been controlled by the rise of monocytes number²⁶. According to our results, treatment with *Portulaca oleracea* exerts an anti-inflammatory effect in HFFD rats, which is similar with many studies, Lee et al. shows by an experiment in mice that the *P. oleracea* suppresses inflammation at the vascular level²⁷, Xiaohang et al reveals that *P.oleracea* reduces expression of cytokine suffer from ulcerative colitis²⁸. Many nutritional and pharmacological agents, including polyphenols and flavonoids have been studied to determine their anti-inflammatory effects²⁹. Many studies have shown that flavonoids exhibit their pharmacological activities, including anti-inflammatory, through the inhibition of important regulatory enzymes³⁰. Apigenin, a polyphenol, inhibits the proliferation and apoptosis of monocytes and lymphocytes during leukemia and inhibits the platelet by blocking TxA₂ (TP) receptors³¹. The treatment with *Aquilaria malaccensis* leading to a reduction of inflammatory signs in rats, probably because of the richness of the plant in different secondary metabolites such as polyphenols and flavonoids. In addition agarwood contains bioactive substances such as 2-(2-phenylethyl) chromones and eudesmol, guaiene³², these compounds have identified by their anti-inflammatory and immunomodulatory activities by reducing the toxicity of cytokines³³. Our results show that the level of cardiac MDA was higher in HFFD-fed rats than the control. High fructose-fat diet has been identified to increase oxidative stress in tissues. Free radical causes cell injury via the mechanism including lipid peroxidation that leads to tissue damage³⁴. Excessive ROS production attacks local cell organelles, including membrane lipids, resulting in lipid peroxidation³⁵. The reduction of lipid peroxidation rate after the treatment of *A.malaccensis*, may be due to antioxidant substances presented in the extract, HPLC analysis of crude extracts of *A. malaccensis* reveal the presence

of Apigenin, Epicatechin, Naringenin Quercetin, Kampferol and Rutin. Some these phytochemicals have antioxidant activity where they offer protection against damage and the risk of developing metabolic diseases³⁶. The potent molecule such as flavonoids was present in the bark and also the plant has an important antioxidant value³⁷ and also has a significant activity of scanning of free radicals³⁸.

Conclusion

In conclusion, this study clearly concluded that the Methanol extract of bark *A.malaccensis* and leave *P.oleracea* possess the ability to control the immune cells associated with HFFD, and protective activity on heart cells, which in turn improve cardiovascular systems.

References

1. Basciano H., Federico L. and Adeli K. (2005). Fructose, insulin resistance, and metabolic dyslipidemia. *Nutr.Metab.*, 2, 5. <https://doi:10.1186/1743-7075-2-5>.
2. Derouiche S., Kawther A., Manel D., Soumya B.A. and Kechrid Z. (2013). The effects of copper supplement on zinc status, enzymes of zinc activities and antioxidant status in alloxan-induced diabetic rats fed on zinc over-dose diet. *International Journal of Nutrition and Metabolism*, 5(5), 82-87.
3. Kuller L.H. (1997). Dietary fat and chronic diseases: epidemiologic overview. *J. Am. Diet. Assoc.*, 97(7 Suppl), S9-S15.
4. Rizkalla S.W. (2010). Health implications of fructose consumption: A review of recent data. *Nutr.Metab.*, 7, 82. <https://doi:10.1186/1743-7075-7-82>.
5. Tappy L., Lê K.A., Tran C. and Paquot N. (2010). Fructose and metabolic diseases: new findings, new questions. *Nutrition.*, 26(11-12), 1044-1049. <https://doi:10.1016/j.nut.2010.02.014>.
6. Monzo-Beltran L., Vazquez-Tarragón A. and Cerdà C. (2017). One-year follow-up of clinical, metabolic and oxidative stress profile of morbid obese patients after laparoscopic sleeve gastrectomy. 8-oxo-dG as a clinical marker. *Redox Biol.*, 12, 389-402. <https://doi:10.1016/j.redox.2017.02.003>.
7. Rakotoarivelo N.H., Rakotoarivony F., Ramarosandratana A.V., Jeannoda V.H., Kuhlman A.R., Randrianasolo A. and Bussmann R.W. (2015). Medicinal plants used to treat the most frequent diseases encountered in Ambalabe rural community, Eastern Madagascar. *Journal of ethnobiology and ethnomedicine*, 11(1), 68. doi:10.1186/s13002-015-0050-2.
8. Mubashir H.M., Bahar A., Showkat R.M., Bilal A.Z. and Nahida T. (2011). Portulacaoleracea L. A Review. *J. Pharm. Res.*, 4(9), 3044-3048.
9. Saikia P. and Khan M.L. (2014). Ecological Features of Cultivated Stands of *Aquilaria malaccensis* Lam. (Thymelaeaceae), a Vulnerable Tropical Tree Species in Assamese Homegardens. *Int. J. For. Res.*, 2014, Article ID 140926, 16. <http://dx.doi.org/10.1155/2014/140926>.
10. Hashim Y.Z., Kerr P.G., Abbas P. and Mohd Salleh H. (2016). *Aquilaria* spp. (agarwood) as source of health beneficial compounds: A review of traditional use, phytochemistry and pharmacology. *J. Ethnopharmacol.*, 189, 331-360.
11. Tobey T.A., Mondon C.E., Zavaroni I. and Reaven G.M. (1982). Mechanism of insulin resistance in fructose-fed rats. *Metabolism*, 31(6), 608-612.
12. Meng R., Da-Long Z., Yan B., Dong-Hui Y. and Ya-Ping W. (2011). Anti-Oxidative Effect of Apocynin on Insulin Resistance in High-Fat Diet Mice. *Ann. Clin. Lab. Sci.*, 41(3), 236-243.
13. Sastre J., Pallardó F.V., García de la Asunción J. and Viña J. (2000). Mitochondria, oxidative stress and aging. *Free Radic. Res.*, 32(3), 189-198.
14. Bradford M.M. (1976). Rapid and sensitive method for the quantitation of microgram quantities of protein utilizing a principle of protein-dye binding. *Anal. Biochem.*, 72, 248-254.
15. Shankar S., Kumar D. and Srivastava R.K. (2013). Epigenetic Modifications by Dietary Phytochemicals: Implications for Personalized Nutrition. *Pharmacol. Ther.*, 138(1), 1-17.
16. Corbi G., Conti V., Davinelli S., Scapagnini G., Filippelli A. and Ferrara N. (2016). Dietary Phytochemicals in Neuroimmunoaging: A New Therapeutic Possibility for Humans. *Front Pharmacol.*, 7, 364. <https://doi:10.3389/fphar.2016.00364>.
17. Fontaine K.R., Redden D.T., Wang C.A., Westfall O. and Allison D.B. (2003). Years of life lost due to obesity. *JAMA.*, 289(2), 187-193.
18. Oliva L., Aranda T., Caviola G., Fernández-Bernal A., Alemany M., Fernández-López J.A. and Remesar X. (2017). In rats fed high-energy diets, taste, rather than fat content, is the key factor increasing food intake: a comparison of a cafeteria and a lipid-supplemented standard diet. *Peer J.*, 5, e3697. <https://doi.org/10.7717/peerj.3697>.
19. Hasani-Ranjbar S., Jouyandeh Z. and Abdollahi M. (2013). A systematic review of anti-obesity medicinal plants - an update. *J. Diabetes Metab. Disord.*, 12(1), 28. <https://doi:10.1186/2251-6581-12-28>.
20. Zheng J., Zheng S., Feng Q., Zhang Q. and Xiao X. (2017). Dietary capsaicin and its anti-obesity potency: from mechanism to clinical implications. *Bioscience reports*, 37(3), BSR20170286. <https://doi:10.1042/BSR20170286>.

21. Gomez-Smith M., Karthikeyan S., Jeffers M.S., Janik R., Thomason L.A., Stefanovic B. and Corbett D. (2016). A physiological characterization of the Cafeteria diet model of metabolic syndrome in the rat. *Physiol. Behav.*, 167, 382-391.
22. Pektas M.B., Koca H.B., Sadi G. and Akar F. (2016). Dietary Fructose Activates Insulin Signaling and Inflammation in Adipose Tissue: Modulatory Role of Resveratrol. *Biomed Res. Int.*, 2016, 8014252. <https://doi.org/10.1155/2016/8014252>.
23. Bratoeva K., Stoyanov G.S., Merdzhanova A. and Radanova M. (2017). Manifestations of Renal Impairment in Fructose-induced Metabolic Syndrome. *Cureus.*, 9(11), e1826. <https://doi.org/10.7759/cureus.1826>.
24. Klein A.V. and Kiat H. (2015). The mechanisms underlying fructose-induced hypertension: a review. *J. Hypertens.*, 33(5), 912-920.
25. Atoussi N., Guediri S. and Derouiche S. (2018). Changes in Haematological, Biochemical and Serum Electrolytes Markers in Women Breast Cancer Patients. *SJRAB.*, 3(2), 173-177.
26. Anfal D. and Samir D. (2017). Study of fluoride-induced haematological alterations and liver oxidative stress in rats. *World J Pharm Pharmsci*, 6(5), 211-221.
27. Lee A.S., Lee Y.J., Lee S.M., Yoon J.J., Kim J.S., Kang D.G. and Lee H.S. (2012). Portulaca oleracea ameliorates diabetic vascular inflammation and endothelial dysfunction in db/db mice. *Evidence-Based Complementary and Alternative Medicine*, 2012. Article ID 741824, 9. <https://doi.org/10.1155/2012/741824>.
28. Yang X., Yan Y., Li J., Tang Z., Sun J., Zhang H. and Liu L. (2016). Protective effects of ethanol extract from Portulacaoleracea L on dextran sulphate sodium-induced mice ulcerative colitis involving anti-inflammatory and antioxidant. *Am. J. Transl. Res.*, 8(5), 2138-2148.
29. Cialdella-Kam L., Nieman D., Knab A., Shanely R., Meaney M., Jin F. and Ghosh S. (2016). A Mixed Flavonoid-Fish Oil Supplement Induces Immune-Enhancing and Anti-Inflammatory Transcriptomic Changes in Adult Obese and Overweight Women-A Randomized Controlled Trial. *Nutrients.*, 8(5), 277. <https://doi.org/10.3390/nu8050277>.
30. Panche A.N., Diwan A.D. and Chandra S.R. (2016). Flavonoids: an overview. *J. Nutr. Sci.*, 5, e47. <https://doi.org/10.1017/jns.2016.41>.
31. Shukla S. and Gupta S. (2010). Apigenin: A Promising Molecule for Cancer Prevention. *Pharm. Res.*, 27(6), 962-978. <https://doi.org/10.1007/s11095-010-0089-7>.
32. Konishi T., Konoshima T., Shimada Y. and Kiyosawa S. (2002). Six new 2-(2-phenylethyl) chromones from Agarwood. *Chem. Pharm. Bull. (Tokyo).*, 50(3), 419-422.
33. Yadav D.K., Mudgal V., Agrawal J., Maurya A.K., Bawankule D.U., Chanotiya C.S, Khan F. and Thul S.T. (2013). Molecular docking and ADME studies of natural compounds of Agarwood oil for topical anti-inflammatory activity. *Curr.Comput. Aided Drug Des.*, 9(3), 360-370.
34. Derouiche S., Zeghibe K., Gharbi S. and Khelef Y. (2017). In-vivo study of stress oxidative and liver damage in rats exposed to acetate lead. *Int. Res. J. Biological Sci.* 6(9), 1-6.
35. Tangvarasittichai S. (2015). Oxidative stress, insulin resistance, dyslipidemia and type 2 diabetesmellitus. *World J. Diabetes*, 6(3), 456-480. <https://doi.org/10.4239/wjd.v6.i3.456>.
36. Lee M.T., Lin W.C., Yu B. and Lee T.T. (2017). Antioxidant capacity of phytochemicals and their potential effects on oxidative status in animals-A review. *Asian-Australas J. Anim. Sci.*, 30(3), 299-308. <https://doi.org/10.5713/ajas.16.0438>.
37. Khoo H.E., Azlan A., Kong K.W. and Ismail A. (2016). Phytochemicals and Medicinal Properties of Indigenous Tropical Fruits with Potential for Commercial Development. *Evid Based Complement Alternat.Med.*, Article ID 7591951, 20 <http://dx.doi.org/10.1155/2016/7591951>.
38. Papalia T., Barreca D. and Panuccio M.R. (2017). Assessment of Antioxidant and Cytoprotective Potential of Jatropha (Jatropha curcas) Grown in Southern Italy. *Int. J. Mol. Sci.*, 18(3), 660. <http://doi.org/10.3390/ijms18030660>.